



小鼠 Treg 调节性 T 细胞检测试剂盒

Anti-Mouse Treg Flow Kit (FOXP3 PE/CD4 FITC/CD25 APC)

Catalog No.: BFK002

Size: 25T

Materials Provided:

Cat No.	Product	Size	Storage
BFK002-1	FITC anti-mouse CD4	250ul	2°C and 8°C. Do not freeze. (12months)
BFK002-2	FITC rat IgG2b Isotype Control	100ul (10T)	2°C and 8°C. Do not freeze. (12months)
BFK002-3	APC anti-mouse CD25	250ul	2°C and 8°C. Do not freeze. (12months)
BFK002-4	APC rat IgG2b K Isotype Control	100ul (10T)	2°C and 8°C. Do not freeze. (12months)
BFK002-5	PE Anti-mouse FOXP3	250ul	2°C and 8°C. Do not freeze. (12months)
BFK002-6	PE mouse IgG1 K Isotype Control	100ul (10T)	2°C and 8°C. Do not freeze. (12months)
BFK002-7	Foxp3/Transcription Factor Fixation/Permeabilization Buffer Set	A buffer: 2x25ml B buffer: 25ml C buffer (10X): 31.5ml	RT, 6months
BFK002-8	Cell Staining Buffer	10ml	2°C and 8°C. Do not freeze. (12months)

Description:

T regulatory (Treg) cells are a subset of T lymphocytes which is characterized by CD4⁺/CD25⁺/FOXP3⁺. These naturally occurring Treg cells originate in the thymus, and comprise 2-10% of peripheral CD4⁺ T cells. It has been shown that Treg cells are able to inhibit T cells proliferation and cytokine production and play critical roles in preventing autoimmunity as well as in controlling tumor immunity and transplantation tolerance. Impaired Treg function or Treg cell deficiency will develop variety of autoimmune diseases, while higher frequency of Treg cells will cause hypo-immune response to pathogens.

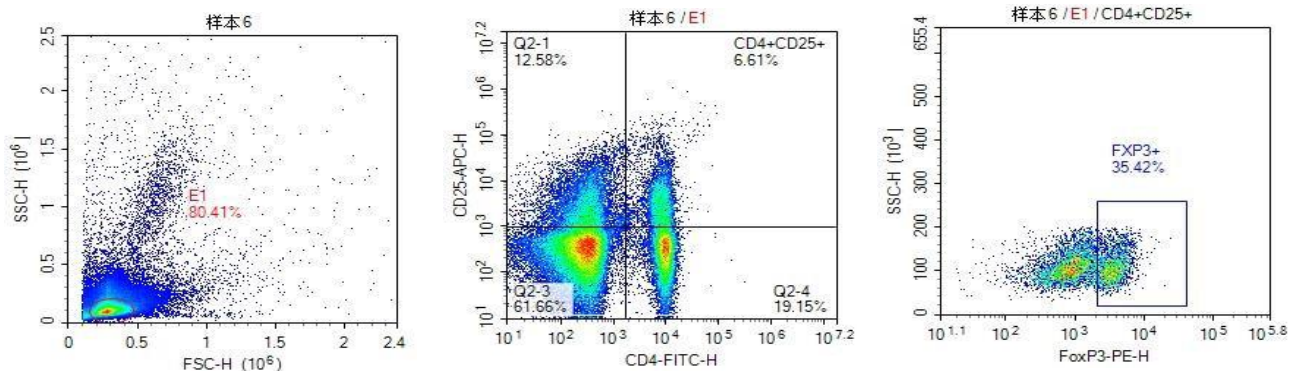
Experimental Methods:

(The following is a general protocol, optimization is required for best results of your experiment. Please contact us by email or by phone for further technical support.)

1. The C buffer(10X) must be freshly diluted to 1X working solution with ddH₂O prior to staining procedures. e.g. Dilute one (1) part C buffer (10X) to nine (9) parts ddH₂O;
2. Take 100μl peripheral blood anticoagulated by EDTA and add to the bottom of 5ml tube;
3. Add 10μl FITC anti-mouse CD4 and 10μl APC anti-mouse CD25 antibody to the bottom of flow tube mixing with the peripheral blood, incubate for 20 minutes at room temperature away from light;
4. Add 2ml A buffer mixing, incubate for 10 minutes at room temperature away from light;
5. Sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;

6. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
7. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
8. Add 1ml B buffer mixing, incubate for 60 minutes at room temperature away from light;
9. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
10. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
11. Add 50ul Cell Staining Buffer
12. Add 10μl PE anti-mouse foxP3 mixing, incubate for 60 minutes at room temperature away from light;
13. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
14. Add 2ml C buffer mixing, sample tube is set to 400g centrifugation for 5 minutes, discard the supernatant;
15. Add 0.5ml C buffer then analyze with flow cytometer with appropriate instrument setting.

Experimental Data :



BALB/c splenocytes were surfaced stained with FITC anti-human CD4 (Cat#BFK002-01) and APC anti-mouse CD25 (Cat#BFK002-03) simultaneously. Cells were fixed and permeabilized followed by intracellular staining with PE anti- mouse FOXP3 (Cat#BFK002-05) .Flow Cytometry was performed using a NovoCyt system.